



**A&A BIOTECHNOLOGY**  
innovating life science

# ExTerminator 96-well

Nucleotide dye terminators removal kit for DNA cycle sequencing reaction samples.

version 1117

192 isolations, 384 isolations

Cat. # 444-192, 444-384

Maximum binding capacity of the minicolumn is 10 µg.

For R&D use only

## Kit Contents

Component	192 isolations	384 isolations	Store at
O purification plate	2 pcs	4 pcs	Room Temp.
R receiving plate	2 pcs	4 pcs	Room Temp.
E elution plate	2 pcs	4 pcs	Room Temp.
Reservoir	1 pcs	1 pcs	Room Temp.
WP bind/wash solution	70 ml	130 ml	Room Temp.
Mix Blue (8-tube strip)	1600 µl (2 pcs)	3200 µl (4 pcs)	Room Temp.
Sterile water (nuclease free, DEPC treated)	15 ml	30 ml	from -20 °C to +20 °C

## Equipment and materials necessary for nucleotide terminator removal that are not included in kit

1. Cycle sequencing reaction mixture
2. Centrifuge with swing-out rotor for plates 5.7 cm high

### NOTE:

Before you start working, we recommend cleaning the work surface using LabZAP™ product (cat. # 040-500)

A&A Biotechnology provides one year guarantee on this kit

The company does not guarantee correct performance of this kit in the event of:

- not adhering to the supplied protocol
- not recommended use of equipment and materials
- the use of other reagents than recommended or which are not a component of the kit
- the use of expired or improperly stored reagents and plates

## Terminators removal protocol

1. Assemble **O** purification plate with **R** receiving plate.
2. Add **5 µl** of **Mix Blue** to cycle sequencing mixture (performed in 10–20 µl).

Note: If cycle sequencing reaction is less than 10 µl add an appropriate volume of sterile water to reach the final volume of 10 µl.

3. Add **100 µl** of **WP** bind/wash solution

Mix by pipeting.

4. Apply samples onto corresponding well of **O** purification plate.
5. Transfer the assembled plates (**O** purification with **R** receiving plates) to swing-out rotor.

Centrifuge for **1 min** at **2000 x g**.

Note: light blue colour of the minicolumn membrane is a result of efficient precipitation of sequencing products.

6. Apply **200 µl** of **WP** bind/wash solution onto each well of **O** purification plate.
7. Centrifuge for **10 min** at **2000 x g**.

8. Carefully separate **O** purification plate from **R** receiving plate.  
Assemble **O** purification plate with **E** elution plate.

9. Apply **35 µl** of **sterile water** directly onto each well of **O** purification plate.

While applying water onto the well be sure that liquid is applied directly onto the resin. If some water stay on the wall the elution will be less effective.

10. Incubate for **2 min** at **room temp**.

11. Transfer the assembled plates (O purification with E elution plates) to swing-out rotor.

Centrifuge for 2 min at 2000 x g.

12. Clear light blue appearance of the eluted samples confirms the correct isolation of cycle sequencing DNA products.  
Blue colour of the sample does not affect the readout of the DNA sequence.  
The samples are ready for thermal denaturation.
13. Store the samples at -20 °C.

## Safety Information



### DANGER

#### WP bind/wash solution

H225 Highly flammable liquid and vapour.

H319 Causes serious eye irritation.

H336 May cause drowsiness or dizziness.

P210 Keep away from heat, sparks, open flames, hot surfaces. No smoking.

P261 Avoid breathing vapours.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.